## COMMENT

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# Immunosenescence in *Caenorhabditis elegans*

Sieun S. Kim<sup>†</sup>, Jooyeon Sohn<sup>†</sup> and Seung-Jae V. Lee<sup>\*</sup>

## Abstract

Immunosenescence is an age-dependent decline in immune functions and hallmark of aging in diverse species, ranging from invertebrates to mammals. However, identifying the factors responsible for immunosenescence is challenging because of the complexity of immune systems and aging in mammals. The roundworm *Caenorhabditis elegans* is suitable for understanding immunosenescence because of its simple immune system and rapid aging process. In this review, we discuss the advances in our understanding of immunosenescence in *C. elegans*. PMK-1/p38 mitogen-activated protein kinase (MAPK), SKN-1/NRF, and ZIP-10/bZIP transcription factor regulate immunosenescence through p38 MAPK and insulin/IGF-1 signaling pathways. Because these factors and pathways are evolutionarily conserved, the findings discussed in this review may help understand the mechanisms underlying immunosenescence and develop new treatment therapy for immunosenescence in humans.

Keywords: Immunosenescence, Caenorhabditis elegans, Aging, MAPK, SKN-1/NRF, Insulin/IGF-1

#### Background

Aging has been regarded as a universal and inevitable biological process in almost all living organisms. One of the hallmarks of aging is immunosenescence, a gradual age-dependent decline in immune functions. Although the contexts of immunosenescence in humans are controversial [1, 2], decreases in the efficiency of adaptive immunity and innate immunity, and in immune responses to vaccination have been reported [3, 4, 5]. For example, the elderly are vulnerable to infection with novel pathogens, such as SARS-CoV-2, although the protection effect of vaccine on older adults are the same as that on younger people [2, 6]. Several recent studies have helped us understand the molecular mechanisms driving immunosenescence and develop strategies to delay or even reverse immunosenescence. Although the hallmarks of immunosenescence have been proposed [5, 7],

<sup>†</sup>Sieun S. Kim and Jooyeon Sohn contributed equally to this work.

\*Correspondence: seungjaevlee@kaist.ac.kr

Department of Biological Sciences, Korea Advanced Institute of Science and Technology, 291 Daehak-ro, Yuseong-gu, Daejeon 34141, South Korea

the causal factors for immunosenescence remain poorly understood.

Because of its short lifespan and genetic tractability, the roundworm Caenorhabditis elegans is an excellent model for studying basic animal physiology, including aging, stress response, immunity, and immunosenescence [8, 9, 10]. Various differences and similarities exist between immunity and immunosenescence in *C. elegans* and those in mammals. C. elegans lacks an adaptive immune system, specialized immune cells, and NF-ĸB signaling, which are crucial for mammalian immunity [10, 11]. However, C. elegans has an innate immune system regulated by evolutionarily conserved signaling pathways, including p38 MAPK, insulin/IGF-1 signaling (IIS) and DBL-1/TGF-  $\beta$  pathways [10, 11, 12, 13, 14, 15]. C. elegans displays an age-dependent decline in immunity, as demonstrated by using Pseudomonas aeruginosa, opportunistic pathogenic bacteria to humans, as a model pathogen [16, 17, 18, 19]. The majority of mammalian immunosenescence research has focused on the aging of specialized immune cells. In contrast, studies on C. elegans immunosenescence have mostly reported agedependent impairments in immune organ functions and



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signaling pathways. Nevertheless, the immune systems in mammals are very complex, and therefore studies using *C. elegans* can provide insights into fundamental mechanisms underlying immunosenescence.

In this short review, we discuss findings regarding immunosenescence in *C. elegans*, focusing on three papers that functionally characterized the genetic factors that contribute to immunosenescence [16, 17, 19]. These papers demonstrated that PMK-1/p38 mitogen-activated protein kinase (MAPK), SKN-1/NRF, and DAF-2/insulin/ IGF-1 receptor, established regulators of aging, immunity, and stress responses [12, 14, 17, 20, 21, 22], affect immunosenescence via acting in the p38 MAPK and IIS pathways. Because these signaling pathways are evolutionarily conserved from *C. elegans* to mammals, the findings we discuss here will lead to the development of therapeutic strategies for delaying or even reversing immunosenescence in humans.

#### Main text

# Age-dependent decline in p38 MAPK signaling contributes to immunosenescence in *C. elegans*

Mammalian p38 MAPK is crucial for immunity and inflammatory responses [23, 24]. PMK-1, a *C. elegans* 

ortholog of p38 MAPK, has been established as a key regulator of innate immunity, which upregulates transcription factors, including ATF-7/activating transcription factor 7 and SKN-1/NRF [12, 17, 20]. Therefore, investigating the role of PMK-1/p38 MAPK in immunosenescence was an intriguing and vital question to address.

Youngman et al. reported that the age-dependent decline of PMK-1 activity underlies immunosenescence by accelerating the deterioration of the intestine, a main immune organ in C. elegans [16]. The authors showed that PMK-1/p38 MAPK is downregulated during aging at the transcriptional and translational levels. The mRNA levels of PMK-1/p38 MAPK targets decrease with age. The authors then performed survival assays by infecting animals with P. aeruginosa PA14 strain at the juvenile L4 larval stage, and day 3 (young), day 6 (middle age), and day 9 (old) of adulthoods, and determined the role of PMK-1/p38 MAPK in immunosenescence. The effects of pmk-1 mutations on impaired pathogen resistance decrease in aged animals depleted of PMK-1. Therefore, the authors concluded that the reduced levels of PMK-1/p38 MAPK in aged C. elegans contribute to age-dependent impairment of immune responses (Fig. 1A). The authors then showed



transcription factor, resulting in further reduction of DAF-2 signaling, and reverses immunosenescence

that *pmk-1* mutations induce the accumulation of bacteria in the intestine, leading to intestinal damage. In conclusion, gradual downregulation of PMK-1/p38 MAPK promotes intestinal aging and impairs immune function, accumulating pathogenic bacteria in the gut, which induces damage in the intestine. This study highlights the role of PMK-1/p38 MAPK, which protects animals from intestinal tissue damage in young age, and its age-dependent downregulation leads to immunosenescence via a vicious circle of damage and pathogen infection in the intestine.

# SKN-1/NRF prevents immunosenescence in young C. elegans

The bacterial virulence factors, such as bacterial toxins, often induce damage in the host intestine by counteracting the host antimicrobial defense mechanisms, and several stress regulators, including the components in IIS, regulate immunity [25, 26]. The transcription factor SKN-1/NRF, which regulates oxidative and xenobiotic stress responses, longevity, and immunity, acts in both p38 MAPK and IIS pathways [14, 21, 27]. However, the role of SKN-1/NRF in immunosenescence was unknown.

Papp et al. reported that SKN-1/NRF is required for normal immunity against gram-negative P. aeruginosa PA14 and gram-positive *Enterococcus faecalis* [17]. PA14 infection increases the nuclear localization of SKN-1/ NRF through Toll/IL-1 Receptor domain protein/TIR-1 and PMK-1/p38 MAPK and subsequently increases the expression of SKN-1/NRF target genes, y glutamylcysteine synthetase 1 (gcs-1) and glutathione S-transferase 4 (gst-4). They also showed an age-dependent decrease in the expression of SKN-1/NRF target genes upon PA14 infection. The authors experimentally evaluated the relationship between SKN-1/NRF activity and immunosenescence and found that SKN-1/NRF target gene expression decreases with age (Fig. 1A). In addition, the survival of day 4 wild-type animals on PA14 is similar to that of day 1 skn-1 mutant animals. These results indicate that a decline in SKN-1/NRF function contributes to immunosenescence. Furthermore, SKN-1/NRF is required to enhance pathogen resistance mediated by reduced IIS or oxidative preconditioning. However, the hyperactivation of SKN-1/NRF by the genetic inhibition of WDR-23, a direct repressor of SKN-1, impairs pathogen resistance. Thus, the optimal activity of SKN-1/ NRF appears crucial for maintaining proper immune responses, consistent with our recent study on the proper regulation of SKN-1/NRF for enhanced immunity and healthy longevity [28]. Overall, this study identifies SKN-1/NRF as a critical factor that contributes to preventing premature immunosenescence in C. elegans.

# Reduction of IIS can reverse immunosenescence in *C. elegans* through a positive feedback loop

Increased innate immunity at a young age is a general feature in various long-lived *C. elegans* mutants [13, 14, 25, 29, 30, 31]. Thus, interventions that alter longevity may have causal effects on immunosenescence, but whether and how these genetic factors affect immunosenescence remained elusive.

Our study showed that DAF-2/insulin/IGF-1 receptor contributes to immunosenescence by inducing INS-7/ insulin-like peptide 7 through DAF-16/FOXO, HSF-1/ heat-shock transcription factor 1, and ZIP-10/bZIP transcription factors [19]. We aimed to identify longevity-promoting interventions that modulate immunosenescence by using five long-lived mutants: insulin/ IGF-1 receptor-defective daf-2, sensory neuron-compromised osm-5, dietary restricted eat-2, mitochondrial respiration-impaired isp-1, and germline-deficient glp-1 mutants. Surprisingly, we found that day 9 (old) daf-2 mutants displayed enhanced pathogen resistance against PA14 compared to day 1 (young) daf-2 mutants that are more resistant to PA14 than wild-type animals. The effect is, at least in part, independent of PA14 intake or clearance. These results indicate that genetic inhibition of DAF-2, which substantially increases lifespan, stress resistance and immunity [13, 14, 32, 33, 34], can cause immune rejuvenation (Fig. 1 B).

DAF-16/FOXO, SKN-1/NRF, and HSF-1 are key transcription factors acting downstream of the DAF-2/insulin/IGF-1 receptor [13, 14], and are crucial for longevity and immunity [17, 25, 35, 36]. We showed that DAF-16/ FOXO and HSF-1 are required for enhanced immunocompetence in old *daf-2* mutants compared with young *daf-2* mutants, whereas SKN-1/NRF is not essential. We also found that depletion of PMK-1/p38 MAPK, a major immune-regulatory factor whose downregulation contributes to immunosenescence [16], is not required for the further enhanced immunocompetence of aged *daf-2* mutants. Thus, DAF-16/FOXO and HSF-1 are key factors that mediate immune rejuvenation in *daf-2* mutants.

We searched for genes that mediate enhanced immunocompetence in old daf-2 mutants by analyzing RNAseq data using day 1 (young) and 9 (old) adult wild-type and daf-2 mutant animals. We found that *ins*-7, an agonistic insulin-like peptide [37, 38, 39], is downregulated during aging in daf-2 mutants by activating DAF-16/ FOXO and HSF-1. Mutations in *ins*-7 are sufficient to enhance pathogen resistance in old animals. We concluded that the downregulation of *ins*-7 contributes to delayed immunosenescence in daf-2 mutants.

We then focused on the ZIP-10/bZIP transcription factor, whose expression increases with age in wild-type animals but decreases in *daf-2* mutants, similar to that

of *ins-7*. ZIP-10/bZIP transcription factor plays a role in the defense response against pathogens in plants [40]. We showed that genetic inhibition of ZIP-10 suppresses age-dependent upregulation of *ins-7* and increases PA14 resistance in day 9 adult animals. Therefore, downregulation of ZIP-10/bZIP transcription factor in aged *daf-2* mutants prevents immunosenescence by increasing the activities of DAF-16/FOXO and HSF-1 by decreasing the levels of INS-7 (Fig. 1 B). This study provides clues for reversing immunosenescence by revealing the role of an immunity-regulating positive feedback loop in IIS in aged *C. elegans*.

#### Conclusions

Many previous studies have suggested that age-dependent accumulation of PA14 along with the extensive deterioration of the intestine, a critical immune organ in C. elegans and mammals, is the primary cause of immunosenescence [12, 41, 42, 43]. Consistent with the reports, the two studies discussed here have demonstrated that agedependent decline in critical immune factors, PMK-1/ p38 MAPK and SKN-1/NRF, in the intestine, are responsible for immunosenescence [16, 17]. While increased colonization of PA14 in the intestine in aged C. elegans is the main cause of immunosenescence, our work suggests that genetic inhibition of *daf-2* can further increase immunocompetence in old age, even under conditions with PA14 accumulation [19]. The abovementioned data highlight the complexity of the links between immunity and aging, suggesting that the relationship is context dependent.

Because studies regarding immunosenescence in *C. elegans* are limited, identifying novel factors that delay or reverse immune aging is required to further understand the relationship between immunity and aging. In *C. elegans*, several signaling pathways, including autophagy, unfolded protein response, and DBL-1/TGF-  $\beta$  pathway, regulate immunity and aging, in addition to p38 MAPK and IIS pathways that are discussed here [10, 11, 15, 44]. Therefore, studying the roles of all these pathways in the context of age-dependent changes in immunity will provide additional mechanistic insights into immunosenescence. In addition, as immunosenescence studies in *C. elegans* have been focused on antibacterial immunity, research regarding age-dependent changes in immune responses against virus and fungi will be crucial.

The studies discussed here identified and characterized genetic factors that regulate immunosenescence in *C. elegans.* In addition, our study provides information for potential strategies to reverse immune aging [19]. As many factors that regulate aging and immunity are evolutionarily conserved from *C. elegans* to mammals, factors and pathways discussed here may also regulate immunosenescence in humans. Consistent with this possibility, p38 MAPK and insulin/IGF-1 signaling contributes to immunosenescence in mammals [5, 45]. Thus, further research is required to unravel the molecular mechanism of immunosenescence in humans to improve treatment therapy of age-related pathologies.

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#### Authors' contributions

S.S.K., J.S., and S-J.V.L. wrote the manuscript. All authors read and approved the manuscript.

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#### Declarations

## Ethics approval and consent to participate

None required.

#### **Consent for publication**

All authors have approved the paper for publication.

#### **Competing interests**

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#### References

- 1. Pawelec G, et al. The conundrum of human immune system "senescence". Mech Ageing Dev. 2020;192:111357.
- Fulop T, et al. Immunosenescence and altered vaccine efficiency in older subjects: a myth difficult to change. Vaccines. 2022;10(4):607.
- Panda A, et al. Human innate immunosenescence: causes and consequences for immunity in old age. Trends Immunol. 2009;30(7):325–33.
- Franceschi C, et al. Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. Mech Ageing Dev. 2007;128(1):92–105.
- Aiello A, et al. Immunosenescence and its hallmarks: how to oppose aging strategically? A review of potential options for therapeutic intervention. Front Immunol. 2019;10:2247.
- Crooke SN, et al. Immunosenescence and human vaccine immune responses. Immun Ageing. 2019;16:25.
- Rodrigues LP, et al. Hallmarks of aging and immunosenescence: connecting the dots. Cytokine Growth Factor Rev. 2021;59:9–21.
- Park HH, Jung Y, Lee SV. Survival assays using *Caenorhabditis elegans*. Mol Cells. 2017;40(2):90–9.
- Kwon S, Kim EJE, Lee SV. Mitochondria-mediated defense mechanisms against pathogens in *Caenorhabditis elegans*. BMB Rep. 2018;51(6):274–9.
- 10. Fabian DK, et al. Functional conservation in genes and pathways linking ageing and immunity. Immun Ageing. 2021;18(1):23.
- 11. Engelmann I, Pujol N. Innate immunity in *C. elegans*. Adv Exp Med Biol. 2010;708:105–21.
- 12. Kim DH, et al. A conserved p38 MAP kinase pathway in *Caenorhabditis* elegans innate immunity. Science. 2002;297(5581):623–6.
- Lee Y, et al. Genes and pathways that influence longevity in *Caenorhabditis elegans*. In: Mori N, Mook-Jung I, editors. Aging mechanisms: longevity, metabolism, and brain aging. Tokyo: Springer Japan; 2015. p. 123–69.
- 14. An SWA, et al. Longevity regulation by insulin/IGF-1 Signalling, in ageing: lessons from *C. elegans.* 2017:63–81.

- Kim DH, Ewbank JJ. Signaling in the innate immune response. Worm-Book. 2018;2018:1–35.
- Youngman MJ, Rogers ZN, Kim DH. A decline in p38 MAPK signaling underlies immunosenescence in *Caenorhabditis elegans*. PLoS Genet. 2011;7(5):e1002082.
- Papp D, Csermely, Soti C. A role for SKN-1/Nrf in pathogen resistance and immunosenescence in *Caenorhabditis elegans*. PLoS Pathog. 2012;8(4):e1002673.
- McHugh DR, et al. DAF-16 and SMK-1 Contribute to Innate Immunity During Adulthood in *Caenorhabditis elegans*. G3 (Bethesda). 2020;10(5):1521–39.
- Lee Y, et al. Reduced insulin/IGF1 signaling prevents immune aging via ZIP-10/bZIP-mediated feedforward loo. J Cell Biol. 2021;220(5).
- Shivers RP, et al. Phosphorylation of the conserved transcription factor ATF-7 by PMK-1 p38 MAPK regulates innate immunity in *Caenorhabditis elegans*. PLoS Genet. 2010;6(4):e1000892.
- Blackwell TK, et al. SKN-1/Nrf, stress responses, and aging in *Caenorhabdi*tis elegans. Free Radic Biol Med. 2015;88(Pt B):290–301.
- 22. Andrusiak MG, Jin Y. Context specificity of stress-activated mitogen-activated protein (MAP) kinase signaling: the story as told by *Caenorhabditis elegans*. J Biol Chem. 2016;291(15):7796–804.
- Cuenda A, Rousseau S. p38 MAP-kinases pathway regulation, function and role in human diseases. Biochim Biophys Acta. 2007;1773(8):1358–75.
- Huang G, Shi LZ, Chi H. Regulation of JNK and p38 MAPK in the immune system: signal integration, propagation and termination. Cytokine. 2009;48(3):161–9.
- Garsin DA, et al. Long-lived C. elegans daf-2 mutants are resistant to bacterial pathogens. Science. 2003;300(5627):1921.
- Altintas O, Park S, Lee SJ. The role of insulin/IGF-1 signaling in the longevity of model invertebrates, *C. elegans* and *D. melanogaster*. BMB Rep. 2016;49(2):81–92.
- McCallum KC, Garsin DA. The role of reactive oxygen species in modulating the *Caenorhabditis elegans* immune response. PLoS Pathog. 2016;12(11):e1005923.
- Park H-EH, et al. A PTEN variant uncouples longevity from impaired fitness in *Caenorhabditis elegans* with reduced insulin/IGF-1 signaling. Nat Commun. 2021;12(1):5631.
- Alper S, et al. The Caenorhabditis elegans germ line regulates distinct signaling pathways to control lifespan and innate immunity. J Biol Chem. 2010;285(3):1822–8.
- Yunger E, et al. Innate immunity mediated longevity and longevity induced by germ cell removal converge on the C-type lectin domain protein IRG-7. PLoS Genet. 2017;13(2):e1006577.
- Tiku V, et al. Nucleolar fibrillarin is an evolutionarily conserved regulator of bacterial pathogen resistance. Nat Commun. 2018;9(1):3607.
- Kenyon C, et al. A C. *elegans* mutant that lives twice as long as wild type. Nature. 1993;366(6454):461–4.
- 33. Kenyon CJ. The genetics of ageing. Nature. 2010;464(7288):504-12.
- Murphy CT, Hu PJ. Insulin/insulin-like growth factor signaling in *C. elegans*. WormBook. 2013:1–43.
- Singh V, Aballay A. Heat-shock transcription factor (HSF)-1 pathway required for *Caenorhabditis elegans* immunity. Proc Natl Acad Sci. 2006;103(35):13092–7.
- Miyata S, et al. DAF-16-dependent suppression of immunity during reproduction in *Caenorhabditis elegans*. Genetics. 2008;178(2):903–18.
- 37. Murphy CT, et al. Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*. Nature. 2003;424(6946):277–83.
- Murphy CT, Lee S-J, Kenyon C. Tissue entrainment by feedback regulation of insulin gene expression in the endoderm of *Caenorhabditis elegans*. Proc Natl Acad Sci. 2007;104(48):19046–50.
- Lee SJ, Murphy CT, Kenyon C. Glucose shortens the life span of *C. elegans* by downregulating DAF-16/FOXO activity and aquaporin gene expression. Cell Metab. 2009;10(5):379–91.
- Alves MS, et al. Plant bZIP transcription factors responsive to pathogens: a review. Int J Mol Sci. 2013;14(4):7815–28.
- Schmucker DL, Thoreux K, Owen RL. Aging impairs intestinal immunity. Mech Ageing Dev. 2001;122(13):1397–411.
- Schmucker DL. Intestinal mucosal immunosenescence in rats. Exp Gerontol. 2002;37(2-3):197–203.
- 43. Mallo GV, et al. Inducible antibacterial defense system in *C. elegans.* Curr Biol. 2002;12(14):1209–14.

- Lee GY, Sohn J, Lee SV. Combinatorial approach using *Caenorhabdi*tis elegans and mammalian Systems for Aging Research. Mol Cells. 2021;44(7):425–32.
- Salminen A, Kaarniranta K, Kauppinen A. Insulin/IGF-1 signaling promotes immunosuppression via the STAT3 pathway: impact on the aging process and age-related diseases. Inflamm Res. 2021;70(10-12):1043–61.

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